Fragile X–Associated Tremor/Ataxia Syndrome
Influence of the FMR1 Gene on Motor Fiber Tracts in Males With Normal and Premutation Alleles
Jun Yi Wang, PhD; David Hessl, PhD; Andrea Schneider, PhD; Flora Tassone, PhD; Randi J. Hagerman, MD; Susan M. Rivera, PhD

**IMPORTANCE** Individuals with the fragile X premutation express expanded CGG repeats (repeats 55-200) in the FMR1 gene and elevated FMR1 messenger RNA (mRNA) levels, both of which may underlie the occurrence of the late-onset neurodegenerative disorder fragile X–associated tremor/ataxia syndrome (FXTAS). Because the core feature of FXTAS is motor impairment, determining the influence of FMR1 mRNA levels on structural connectivity of motor fiber tracts is critical for a better understanding of the pathologic features of FXTAS.

**OBJECTIVE** To examine the associations of CGG repeat and FMR1 mRNA with motor-related fiber tracts in males with premutation alleles.

**DESIGN AND SETTING** A case-control study conducted at the University of California, Davis, from April 1, 2008, through August 31, 2009. All data were collected masked to the carrier status of the FMR1 gene.

**PARTICIPANTS** Thirty-six male premutation carriers with FXTAS and 26 male premutation carriers without FXTAS were recruited through their family relationships with children affected by fragile X syndrome. The controls were 34 unaffected family members and healthy volunteers from the local community.

**MAIN OUTCOMES AND MEASURES** The CGG repeat lengths and FMR1 mRNA expression levels in peripheral blood lymphocytes, motor functioning, and white matter structural integrity that were estimated using diffusion tensor imaging. After data collection, we selected 4 motor tracts to reconstruct using diffusion tensor tractography, namely, the middle and superior cerebellar peduncles, descending motor tracts (containing the corticospinal, corticobulbar, and corticopontine tracts), and the anterior body of the corpus callosum.

**RESULTS** All fiber tracts exhibited weaker structural connectivity in the FXTAS group (decreased 5%-53% from controls, \( P < .02 \)). Genetic imaging correlation analysis revealed negative associations of CGG repeat length and FMR1 mRNA with connectivity strength of the superior cerebellar peduncles in both premutation groups (partial \( r^2 = 0.23-0.33, P < .004 \)). In addition, the measurements from the corpus callosum and superior cerebellar peduncles revealed a high correlation with motor functioning in all 3 groups (\( r \) between partial least square predicted and actual test scores = 0.41-0.56, \( P \leq .04 \)).

**CONCLUSIONS AND RELEVANCE** Distinct pathophysiologic processes may underlie the structural impairment of the motor tracts in FXTAS. Although both the corpus callosum and superior cerebellar peduncles were of great importance to motor functioning, only the superior cerebellar peduncles exhibited an association with the elevated RNA levels in the blood of fragile X premutation carriers.

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The fragile X mental retardation 1 (FMR1) gene contains a polymorphic trinucleotide CGG repeat in its 5′ untranslated region. The repeat sizes normally range from 5 to 44 but exceed 200 in full mutation, which effectively silences the gene, causing intellectual disability in fragile X syndrome.1,2 CGG repeats from 55 to 200 are within the premutation range. Although the FMR1 protein levels are only slightly reduced in the premutation alleles, the FMR1 messenger RNA (mRNA) levels are significantly elevated3,4 and thought to cause the neurodegenerative disorder fragile X-associated tremor/ataxia syndrome (FXTAS).3-5 FXTAS is a late-onset neurodegenerative disorder that affects approximately 40% of males with premutation alleles and who are older than 50 years.7 A constellation of clinical features is linked to FXTAS, with progressive intention tremor and cerebellar ataxia presenting as core features.8

Structural magnetic resonance imaging (MRI) has the advantage of revealing disease progress in vivo. Previous studies revealed T2 hyperintensities in the middle cerebellar peduncle (MCP), widespread gray matter atrophy and white matter structural impairment, and, a negative correlation between CGG repeat length and brain volume, MCP packing density, and gray matter density of the dorsomedial frontal lobes.9-14 Given that motor deficits are the core clinical features and the prominent involvement of the cerebellum in the pathophysiologic function of FXTAS, it is critical to examine the RNA toxic effect (ie, RNA toxic gain-of-function due to excess FMR1 mRNA)3,4,15 on motor fiber tracts for a better understanding of the pathologic features of FXTAS. In the current study, we reconstructed motor-related fiber tracts from diffusion tensor imaging (DTI)16 and tested the associations of CGG repeat length and FMR1 mRNA with structural connectivity of these motor tracts in premutation carriers with and without FXTAS.

Methods

Research Participants
We recruited 62 male premutation carriers through their family relationships with children affected by fragile X syndrome from April 1, 2008, through August 18, 2009, before a major scanner upgrade that resulted in incomparable DTIs before and after the upgrade. Unaffected family members, along with healthy males recruited from the local community, served as controls (n = 34). These controls and 26 of the premutation carriers were the same participants in a recent investigation of age-related changes in fragile X premutation.13 We identified premutation alleles using FMR1 DNA testing17 and measured FMR1 mRNA using a quantitative-fluorescence reverse transcription–polymerase chain reaction method.4 In addition, we assessed FXTAS stage with tremor or ataxia intensity,8,18 behavioral self-regulation using the Behavioral Dyscontrol Scale 2,19 and dexterity using the Purdue Pegboard Dexterity Test.20 We diagnosed 36 carriers as being affected by FXTAS, ranging from FXTAS stage 2 (minor tremor or balance problems with no interference in daily living) to 5 (use of a wheelchair on a daily basis). In multiple regression analysis with age as a covariate, we found that although the FXTAS-negative group showed similar motor abilities as the controls, the FXTAS-positive group exhibited deficits in behavioral self-regulation and dexterity (Table 1).

Table 1. Characteristics of the 96 Research Participants

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Healthy Controls</th>
<th>Fragile X–Associated Tremor/Ataxia Syndrome (FXTAS)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>No. of Participants</td>
<td>Mean (SD) [Range]</td>
</tr>
<tr>
<td>Age, y</td>
<td>34</td>
<td>43.6 (18.4) [18-81]</td>
</tr>
<tr>
<td>Age for older participants, y</td>
<td>15</td>
<td>62.3 (8.1) [51-81]</td>
</tr>
<tr>
<td>CGG length</td>
<td>34</td>
<td>28.2 (4.47) [19-42]</td>
</tr>
<tr>
<td>FMR1 mRNA level</td>
<td>34</td>
<td>1.49 (0.25) [1.0-2.0]</td>
</tr>
<tr>
<td>Behavioral tests</td>
<td></td>
<td></td>
</tr>
<tr>
<td>BDS-2</td>
<td>32</td>
<td>22.4 (3.01) [17-27]</td>
</tr>
<tr>
<td>Purdue Pegboard Dexterity Test*</td>
<td>29</td>
<td>39.7 (6.50) [26-57]</td>
</tr>
</tbody>
</table>

Abbreviations: BDS-2, Behavioral Dyscontrol Scale 2; mRNA, messenger RNA.
* Significantly different from the control group (P ≤ .05).
† Significantly different from the FXTAS-negative group (P ≤ .05).
‡ Total test scores for both hands were used.

Standard Protocol Approvals, Registrations, and Patient Consents
All participants signed informed consent forms issued by the institutional review boards at the University of California, Davis.

Neuroimage Acquisition and Processing
We performed neuroimaging from an MRI scanner with an 8-channel head coil (Siemens Trio 3T MRI scanner; Siemens Medical Solutions). We obtained DTIs with 30 gradient directions using a single-shot, diffusion-weighted echo planar imaging sequence in 72 axial sections of 1.9-mm thickness (no gap) with a 243-mm field of view and a 128 × 128 matrix. The diffusion sensitizing gradients were applied at a b of value 700 s/mm². Five additional images with minimum diffusion weighting were also obtained.

Neuroimage Processing
The DTI processing and tractography have been described in detail in a previous study.13 To summarize, we used the FSL software package (http://fsl.fmrib.ox.ac.uk/fsl/fslwiki/; University of Oxford) for performing eddy current and motion correction and skull stripping.21 We conducted DTI tractography in DTIStudio (http://www.mristudio.org; The Johns Hopkins Medical In-
stitute) by applying a multiple regions-of-interest approach. The fractional anisotropy (FA) threshold for fiber tracking was set at 0.18 and the angle threshold at 70°. The tractography methods have been previously tested, and all measurements reached an intraclass correlation coefficient of 0.9 or more. Fiber tracking was performed masked to the status of the participants.

We reconstructed 4 motor-related fiber tracts: (1) the descending motor tract containing the corticospinal, corticopontine, and corticobulbar tracts; (2) the MCP; (3) the superior cerebellar peduncle (SCP); and (4) the anterior body of the corpus callosum (CC; containing primarily the transcallosal fibers for the premotor and supplementary motor areas). Tractography measurements were FA for assessing fiber directionality, mean diffusivity (MD) for packing density, and tract volume for the number of voxels occupied by the reconstructed fiber tract. To account for individual differences in cranial size, total cranial volume was estimated from the T1-weighted MPRAGE images using the SIENAX function from FSL and was used to normalize tract volumes.

Statistical Analysis

We divided the participants into 3 groups: FXTAS-positive, FXTAS-negative, and control groups. We were able to consider not only the effect of having FXTAS but also the potential differential effect of FMR1 premutation alleles and normal alleles on motor fiber tracts. We applied multiple linear regression analyses to compare both premutation groups with the control group and to predict individual tractography measurements using either CGG repeat length or FMR1 mRNA level. Because all premutation carriers with FXTAS were 50 years or older, only older healthy controls (≥50 years) were included in the comparisons of the FXTAS group with the control group. We used age as a covariate for multiple linear regression. We constructed residual plots to identify possible violations of the assumptions in linear regression and repeated analysis using ranks in case of violation. To predict motor functioning from the combination of 12 tractography measurements, we conducted partial least square (PLS) regression, which decomposed the independent and dependent variables simultaneously to find a set of latent variables explaining the maximum covariance. The quality of the prediction was assessed by calculating Pearson product moment correlation coefficient and paired t test between the original and predicted test scores. The importance of individual tractography measurements to the predictions were assessed by the Pearson product moment correlation coefficient and paired t test between the original and predicted test scores. The importance of individual tractography measurements to the predictions were assessed by the Pearson product moment correlation coefficient between the tractography measurements and PLS factors. We cross-validated the PLS results using the leave-one-out procedure in which we took out the participants one by one and predicted their test scores using the tractography measurements from the remaining participants. A separate PLS regression was performed for each group and for each test because of missing test scores for some of the participants. To correct for multiple comparisons, we applied the Benjamini-Hochberg method of false discovery rate (FDR) to all analyses. All statistical analyses were conducted using the Matlab software package (version 7.10.0; The Mathworks Inc).
observed that another zone, from axial sections 42 to 48, had
pared with the older controls after adjusting for age. We also
TAS-positive group indeed had significantly elevated FA com-
 FA value for these sections (eFigure in Supplement). The FX-
intersect with the CC and, consequently, calculated the mean
tions 48 to 56 as the zone in which the descending motor tracts
the CC led to artificially increased FA of the cerebral pedun-
study revealed that degeneration of the body and splenium of
(v = 2,48) df = 2.57). P Value

Table 2. Group Mean (SD) and Comparisons of Tractography Measurements

<table>
<thead>
<tr>
<th>Variable</th>
<th>HCs (n = 34)</th>
<th>FXTAS Negative (n = 26)</th>
<th>FXTAS Positive (n = 36)</th>
<th>P Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Descending motor tracts</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Tract volume, voxels</td>
<td>1287 (354)</td>
<td>1264 (276)</td>
<td>1295 (274)</td>
<td>.79</td>
</tr>
<tr>
<td>Fractional anisotropy</td>
<td>0.46 (0.02)</td>
<td>0.46 (0.02)</td>
<td>0.48 (0.03)</td>
<td>.004</td>
</tr>
<tr>
<td>Mean diffusivity, μm^2/μs</td>
<td>0.82 (0.03)</td>
<td>0.82 (0.04)</td>
<td>0.82 (0.04)</td>
<td>.96</td>
</tr>
<tr>
<td>Middle cerebellar peduncle</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Tract volume, voxels</td>
<td>2931 (543)</td>
<td>2868 (517)</td>
<td>2893 (521)</td>
<td>.67</td>
</tr>
<tr>
<td>Fractional anisotropy</td>
<td>0.50 (0.02)</td>
<td>0.49 (0.02)</td>
<td>0.48 (0.03)</td>
<td>.003</td>
</tr>
<tr>
<td>Mean diffusivity, μm^2/μs</td>
<td>0.74 (0.03)</td>
<td>0.75 (0.03)</td>
<td>0.74 (0.03)</td>
<td>.003</td>
</tr>
<tr>
<td>Super cerebellar peduncle</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Tract volume, voxels</td>
<td>761 (189)</td>
<td>757 (225)</td>
<td>758 (225)</td>
<td>.92</td>
</tr>
<tr>
<td>Fractional anisotropy</td>
<td>0.47 (0.03)</td>
<td>0.47 (0.03)</td>
<td>0.48 (0.03)</td>
<td>.003</td>
</tr>
<tr>
<td>Mean diffusivity, μm^2/μs</td>
<td>0.98 (0.06)</td>
<td>0.99 (0.05)</td>
<td>0.99 (0.05)</td>
<td>.003</td>
</tr>
<tr>
<td>Corpus callosum anterior body</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Tract volume, voxels</td>
<td>1920 (488)</td>
<td>1960 (451)</td>
<td>1962 (451)</td>
<td>.64</td>
</tr>
<tr>
<td>Fractional anisotropy</td>
<td>0.56 (0.03)</td>
<td>0.55 (0.02)</td>
<td>0.54 (0.03)</td>
<td>.51</td>
</tr>
<tr>
<td>Mean diffusivity, μm^2/μs</td>
<td>0.80 (0.05)</td>
<td>0.80 (0.05)</td>
<td>0.80 (0.05)</td>
<td>.002</td>
</tr>
</tbody>
</table>

Abbreviations: FXTAS, fragile X–associated tremor/ataxia syndrome; HCs, healthy controls.

Results

Group Comparisons

The 4 fiber tracts (Figure 1) were reconstructed successfully for all participants, including the 3 carriers at FXTAS stage 5. In group comparisons, 9 tractography measurements survived the 5% FDR (P ≤ .02). When compared with the older control group, the FXTAS-positive group had significantly lower tract volume of all 4 fiber tracts; higher MD of the SCP, MCP, and CC; lower FA of the SCP; and higher FA of the descending motor tract. In contrast, none of the tractography measurements had any group differences between the FXTAS-negative groups and with SCP FA in the FXTAS-positive group only. In addition, FMR1 mRNA had a negative correlation with SCP FA in the FXTAS-positive group (Figure 2) and CC FA in the control group (partial r² = 0.31, P < .001).

Three premutation carriers had extremely high FMR1 mRNA levels (≥3 SDs). We thus repeated multiple linear regression using rank-ordered data. The results were comparable. The only differences were that SCP MD instead of SCP FA significantly correlated with FMR1 mRNA in the FXTAS-positive group, and the correlation between SCP track volume and FMR1 mRNA became significant at an FDR of 5% (P = .007) using rank-ordered data.

Correlation With Motor Functioning

The PLS regression could predict all motor test scores from the tractography measurements for the 3 groups. Therefore, only the results from the leave-one-out cross-validation have been chosen for presentation. Three tests survived the 5% FDR (P ≤ .02), indicating the significant correlation between the actual and predicted motor test scores (Table 3). The tractography measurements predicted behavioral regulation and dexterity for the FXTAS-positive carriers and behavioral regulation for the controls. The tractography measurements predicted behavioral regulation of FXTAS-negative carriers to a lesser degree, with this value passing the 10% FDR (P = .04). Consistent with the PLS results, none of the paired t tests showed
significant differences between the actual and predicted motor scores (Table 3). In the PLS regression, measurements from the CC and SCP made substantial contributions to the predictions of motor tests for all the 3 groups, whereas those from the descending motor tracts and MCP were important to the prediction of dexterity in both the fragile X-associated tremor/ataxia syndrome (FXTAS) negative (partial $r^2_{3,32} = 0.33, P = .004$) and FXTAS-positive (partial $r^2_{3,32} = 0.24, P = .003$) groups. The SCP fractional anisotropy (FA) showed a negative correlation with CGG repeat length in the FXTAS-positive group (partial $r^2_{2,32} = 0.29, P = .001$). The SCP also showed a negative correlation between FA and FMR1 messenger RNA in the FXTAS-positive group (partial $r^2_{2,33} = 0.23, P = .003$).

### Discussion

This study investigated the link among the FMR1 gene, motor-related fiber tracts, and motor functioning in males with premutation and normal alleles. Two motor fiber tracts, the SCP and the anterior body of the CC, had significant associations with the FMR1 gene and motor functioning. Specifically, we found that both increased CGG repeat length and FMR1 mRNA correlated with lower SCP connectivity in the FXTAS-positive and FXTAS-negative groups. We also found that FMR1 mRNA elevation correlated with reduced CC connectivity in the control group. In addition, measurements from the CC and SCP exhibited high correlation with motor functioning in all 3 groups.

Carrying most of the efferent fibers from the cerebellum, the SCP has a critical role in motor functioning and may be a key structure manifesting the RNA toxicity associated with fragile X premutation. The current study demonstrated a negative dose effect of CGG repeat length and FMR1 mRNA on the connectivity strength of SCP in premutation carriers both with and without FXTAS. Our results are consistent with pathophysiologic findings of FXTAS, including marked dropout of...
Purkinje cells, white matter disease throughout the cerebellum, and the presence of intranuclear inclusions in the inferior olivary and dentate nuclei.31,32 Most of the SCP fibers originate in the deep nuclei (dentate and interposed nuclei), which receive the projections from the Purkinje cells, the only cell type carrying the output from the cerebellar cortex.30 It is reasonable to assume that the negative association between the SCP and the expanded CGG repeats is related to the degeneration of both dentate nucleus and Purkinje cells. Further studies are needed to investigate the particular vulnerability of SCP to the CGG expansion and FMR1 mRNA elevation.

Additional major findings are related to the CC. Previous studies of FXTAS have reported thinning,14 FA reduction,12 and high variability of the tractography measurements of the CC, which correlated with carriers’ FXTAS stage.53 The current results extend these findings by revealing a significantly reduced structural connectivity of this fiber tract in the FXTAS-positive group and the relationship between FMR1 mRNA and CC FA in the control group. However, in the premutation carriers, the correlation between FMR1 mRNA and the CC was not significant; in contrast, the CC displayed progressive degeneration with increasing FXTAS stage.13 It is conceivable that the combination of FMR1 mRNA levels remaining stable throughout adulthood33 and the CC connectivity worsening with FXTAS severity contributed to the disappearance of this correlation in the premutation groups.

In addition, the changes in the CC measurements with FXTAS progression might not be linear, potentially making the situation even more complex. The anterior body of the CC had higher FA and lower MD in some of the patients at the early stages of FXTAS compared with the controls, whereas the directions of change reversed at the advanced stage. The early changes in FA and MD were unlikely caused by the crossing-fiber issue in DTI. Although the degeneration of one crossing fiber tract would lead to artificial elevation of FA for the surviving fiber tract, MD would also increase because of the less restricted water diffusion. The pattern of DTI changes at early stages of FXTAS could be explained by axonal swelling because of mitochondrial dysfunction that has been detected in premutation carriers regardless of FXTAS status.34 One consequence of mitochondrial dysfunction is impaired axonal transport, which relies on mitochondria for its energy supply.35,36 This process leads to accumulation of cellular organelles within the axons, consistent with the early DTI representations. At advanced stages of FXTAS, however, we observed decreased FA and increased MD compared with healthy controls in some of the patients, which was consistent with axonal degeneration. Support for these assumptions comes from mitochondrial DNA mutations in which MD reduction in acute lesions and MD elevation in chronic lesions have been reported.36 This complex pattern of DTI changes could also explain why the correlation with FMR1 mRNA was not observed in the FXTAS-negative group. The FXTAS-negative group was composed of premutation carriers with heterogeneous phenotypes. Some members may develop FXTAS later in their lives, whereas others may not. To confirm, longitudinal studies combining neuroimaging and molecular data will be valuable for delineating the dynamic changes of DTI measurements as FXTAS progresses and pathophysiologic mechanisms underlying the DTI changes.

Regarding the MCP, we failed to replicate the quadratic relationship between CGG length and the MCP diffusivity observed when premutation carriers with and without FXTAS were combined in another study.12 The MCP carries efferent fibers from the pontine nucleus to almost all areas of the cerebellar cortex30 and is the structure that has shown T2 hyperintensive signals (the MCP sign) in a subset of FXTAS carriers44; therefore, the MCP is used as one of the criteria for diagnosing FXTAS.8 In the current cohort, the MCP sign was detected in all 26 carriers at stages 3 to 5, 3 of 10 carriers at stage 2, and 1 of 11 carriers at stage 0, whereas none of the controls showed the MCP sign. Consistent with these neurologic observations, significantly elevated MCP MD was detected in the carriers with FXTAS compared with the controls. However, the MCP sign may actually involve both the MCP and SCP. The SCP is a relatively small structure and shares pathways with the MCP in the cerebellar white matter core. Although visual inspection of T2 signal intensity is useful for clinical diagnosis, the SCP cannot be distinguished well in a clinical read. Tractography in DTI has the advantage of obtaining less subjective, quantitative measurements for correlational analyses, as well as having the capability to distinguish the SCP from the MCP. In the current study, the CGG repeat length had a robust correlation with the reconstructed SCP but not with the MCP even when the FXTAS-positive and FXTAS-negative groups were combined. Differences in methods may account for these discrepancies. The previous DTI study32 conducted tract-of-interest and voxel-based analyses that required image normalization to a standard space to define the anatomical location of a fiber tract. The current study performed DTI tractography in participants’ native spaces to localize a fiber tract. In the brains with severe atrophy, the SCP may be best quantified using tractography in participants’ native space to minimize distortion during image normalization.

Another intriguing finding is the relative sparing of the descending motor tracts in contrast to the severe degeneration of the corpus callosum body despite the fact that the fibers from these 2 structures may terminate in the same cortical areas. Further studies are needed to identify neuroprotective factors that prevent descending motor tracts from the RNA toxicity in fragile X premutation.55

The current study is limited by the well-known crossing-fiber issue in the fiber tracking algorithm, which may cause early termination of fiber propagation or absent fiber branches. Small sample sizes, especially in the case of the FXTAS-negative premutation group (n = 26), may provide insufficient powers to detect a real effect. The FMR1 mRNA levels, which have shown high variability among different tissues, were obtained from peripheral blood leukocytes and thus may not completely reflect the correspondent values in the CNS.37 Because many analyses were performed, there is a need to replicate the study in other samples, which, if consistent or largely consistent with the current findings, would make false discovery of an effect much less likely. The association of reduced FMR1 protein seen in some premutation carriers56,58,59 with structural connectivity is an important remaining ques-

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tion that we hope to explore in future studies. Finally, we conducted a separate study to further examine the correlation between FMR1 mRNA and brain measurements (including the anterior body of the CC) in the control group.

In conclusion, we demonstrate the relevance of the motor fiber tracts to FXTAS, finding associations with both RNA toxicity and motor impairment. Because FXTAS is an age-related neurodegenerative disorder that commonly affects male fragile X premutation carriers older than 50 years, clinicians and researchers should target male premutation carriers 40 years or older for early diagnosis, disease monitoring, and potential treatment. Alternations in the motor fiber tracts detected by imaging techniques may prove to be effective biomarkers of early development of FXTAS in future studies.

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Acquisition of data: Wang, Hessl, Schneider, and Tassone.
Analysis and interpretation of data: Wang, Schneider, and Rivera.
Drafting of the manuscript: Wang.
Critical revision of the manuscript for important intellectual content: All authors.
Statistical analysis: Wang.
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Study supervision: Hessl, Hagerman, and Rivera.
Consult for assessments: Schneider.

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REFERENCES