Williams Syndrome

Neuronal Size and Neuronal-Packing Density in Primary Visual Cortex

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Background: Williams syndrome (WMS) is a rare, genetically based syndrome associated with a hemideletion in chromosome 7 (7q11.22-23) and characterized by a unique constellation of somatic, brain, and cognitive features. Individuals with WMS demonstrate an unusual and uneven neuropsychological profile showing cognitive and visual spatial deficits juxtaposed with relative language preservation and excellent facial recognition.

Objectives: A neuroanatomical hypothesis for these behavioral findings suggests predominant involvement of the dorsal portions of the hemispheres relative to the ventral portions, including preferential involvement of peripheral visual field cortical representations over central representation. Predominant involvement of magnocellular visual pathways, as opposed to parvocellular pathways, is also suggested by this hypothesis.

Subjects: We examined primary visual cortical area 17 in the right and left hemispheres in 6 age- and sex-matched autopsy specimens from 3 WMS-affected brains (1 male and 2 females; mean [SD] age, 44 [14] years) and 3 control brains (1 male and 2 females; mean age, 43 [11] years).

Design: Neurons in layers II, III, IVA, I VB, IVCα, IVCβ, V, and VI were measured using an optical dissector method to determine possible differences between WMS-affected and control brains in cell-packing density, neuronal size, and neuronal size distribution.

Results: We found abnormalities in peripheral visual cortex in WMS-affected brains, but not in magnocellular subdivisions. There was a hemisphere by layer IV interaction and a layer IV left hemisphere and diagnosis interaction in cell-packing density. Williams syndrome-affected brains showed increased cell-packing density in left sublayer IVCβ and an excess of small neurons in left layers IVA, IVCα, IVCβ, V, and VI.

Conclusions: Cell measurements differ in peripheral visual cortical fields of WMS, with significantly smaller, more closely packed cells in some layers on the left side. These cell-packing density and neuronal size differences may be related to visuospatial deficits in this population.

Arch Neurol. 2002;59:1461-1467

Williams syndrome (WMS), a mental retardation syndrome, consists of a unique constellation of somatic, brain, and cognitive features, and is associated with a hemideletion in the short arm of chromosome 7 (7q11.22-23). At least 15 genes are associated with this hemideletion, which affects the same set of genes in nearly all clinically identified WMS. However, there are rare individuals with partial deletions with partial phenotypic manifestations of WMS. Approximately 1 in 25000 births exhibit the deletion and accompanying phenotype. Our histometric studies are part of a multidisciplinary project involving cognition, brain morphology (magnetic resonance imaging and functional magnetic resonance imaging), neurophysiology, and molecular genetics.

Specifically, our research has centered on the description of the neuroanatomical phenotype at the cytoarchitectonic, histometric, and histochemical levels for the purpose of linking, on the one hand, brain change to behavior, and, on the other, brain change to the genomic anomaly.

The microanatomical brain research in our laboratory is driven by a general hypothesis derived from the analysis of behaviors exhibited by individuals who have WMS. The WMS neuropsychological profile is an unusual and uneven one, consisting of deficits in processing visuospatial tasks, relative strength in many aspects of language, and a preserved ability to process human faces.

Figure 1 shows the specific deficits in spatial cognition, deficits that are contrasted with comparable individuals with Down syndrome. Individuals who have WMS also demonstrate
an unusual personality characterized by a lack of fear of strangers, highly affective speech, and occasionally, inappropriate friendliness, and often show a great deal of interest in, and sometimes an ability regarding things musical.5,10

The best neuroanatomical fit for many of the behavioral findings seen in WMS seems to be the primary involvement of the dorsal portions of the hemispheres, which in the caudal half of the brain is concerned with representation and processing of visuospatial information11-14 and in the frontal lobes with release and control of behavior.15 By contrast, behaviors associated with the ventral and perisylvian portions of the hemispheres, concerned with most aspects of language,16-18 object properties of visual and other stimuli,19-21 and programs for the performance of various motor behaviors (eg, speech22-23) seem to be at least relatively spared in WMS. Therefore, one part of the research in our laboratory has focused on comparing histometric features between the dorsal and ventral portions of the cerebral hemispheres. As part of a larger histometric study, we report herein a histometric analysis of the visual cortex of WMS. Specifically, we examined primary visual area 17 halfway between the splenium of the corpus callosum and the occipital pole along the calcarine sulcus (Figure 2); assuming normal topography of visual cortex in WMS, this sampled region would represent mostly peripheral fields pathways24-27 and relates more to the dorsal visual pathway.11,28-29

**SUBJECTS AND METHODS**

**SUBJECTS**

We examined the visual cortex in age- and sex-matched autopsy specimens from 3 WMS-affected cases (1 male and 2 females; mean [SD] age, 44.0 [14] years) and from 3 neurologically and psychiatrically healthy control subjects (1 male and 2 females; mean age, 43.3 [11] years; the Harvard Brain Tissue Resource Center, McLean Hospital, Belmont, Mass). There was no information on handedness. The WMS cases had the 7q11.22-23 deletion determined by fluorescent in situ hybridization. The WMS-affected brains (1033 [104] g) were significantly smaller ($P$<.05) than the control brains (1426 [177.8] g).

![Figure 1](https://example.com/figure1.png)

**Figure 1.** Note that both Williams syndrome–affected cases and Down syndrome–affected subjects are poor in spatial cognition, but in contrasting ways. Drawings and block designs of individuals with Williams syndrome tend to focus on the details at the expense of the whole, whereas individuals with Down syndrome tend to show global configuration but may be poor on internal detail. Reprinted with permission from *Trends in Neuroscience.*5

![Figure 2](https://example.com/figure2.png)

**Figure 2.** Blocks of tissue (7 × 5 × 5 cm) were taken from the medial surface of the left and right occipital lobes, dorsal and ventral to the calcarine sulcus (long arrows). sp indicates splenium; L, lateral ventricle; fx, fornix; pi, pineal gland; th, thalamus; ac, anterior commissure; mb, mamillary body; and h, hypothalamus. Reprinted with permission from Duvernoy HM. *The Human Brain, ’91 Edition.* New York, NY: Springer-Verlag Wien, 1991:279.
The primary visual cortex, area 17, was easily identified in Williams syndrome-affected brains compared with control brains. Whereas in layer IVc of the right hemisphere, there was no significant difference between the 2 groups. Asterisk indicates P<.001.

HISTOLOGICAL STUDY

One WMS-affected brain (subject 1) was processed using the Yakovlev whole-brain method of serial histological sections.30 The postmortem brain tissue from the remaining WMS-affected and control brains (subjects 2-6) was processed from left and right hemisphere blocks (7 × 5 × 5 cm), including dorsal and ventral calcarine banks, and sectioned at 30 µm. Every 10th section was stained with cresyl echt violet for Nissl substance.

AREA 17 CELL MEASURES

The primary visual cortex, area 17, was easily identified in WMS-affected and control brains on the calcarine region. Three fields from the pial surface to the gray-white matter junction were selected where the plane of section was perpendicular or near perpendicular to the pial surface and there was no distortion by rippling, tears or other artifacts. All sections were coded so that the examiner was blind to diagnosis and hemisphere. The architectonic appearance of area 17 in WMS-affected brains is indistinguishable from that in controls, so this form of blinding was deemed to be adequate.

Layers II, III, IVA, IVB, IVCα, IVCβ, V, and VI of area 17 were measured in each hemisphere. Neurons were identified by the presence of a clearly visible, single nucleolus, a feature that distinguishes them from glial cells. Cross-sectional neuronal areas and cell packing densities were measured using the modified dissector method and software of Williams and Rakic.31 Using a universal microscope (Carl Zeiss Inc, Thornwood, NY) under ×500 oil magnification, images captured by a camera (Vidicon; Division of Hamamatsu USA, Bridgewater, NJ) were displayed on a monitor (model GVM 1310; Sony, Toronto, Ontario) that was connected to a personal computer workstation (Macintosh Centris 650; Apple Computers, Cupertino, Calif). The counting chamber (95 × 85 × 20 µm, at ×500) was placed within these images. A photoelectric micrometer (Heidenhain MP-25, Heidenhain, Schaumburg, Ill) interfaced to a National Instruments NB-GPIB card (NB-series cards, NB-GPIB IEEE-488.2, National Instruments Corp, Austin, Tex) in the Macintosh recorded movement in the z-axis. The base of the sections was set to a z-axis reading of 0. A red opaque overlay precluded cell counting below the dimensions of the counting box. With the movement of the stage to 5 µm (7.5 µm for subject 1) above the original position of the base of the section, the screen became transparent and the cells visible. The soma of the neurons were traced on a digitizing tablet, whereby neurons touching the top and left side of the screen were omitted. At a stage level of more than 25 µm (27.5 µm for subject 1) above the original position of the base of the section, the screen turned opaque green, which prevented the measurement of any cells above the optical counting box.

STATISTICAL ANALYSIS

Repeated-measures analysis of variance (ANOVA) was used to determine cell-packing density and neuronal size differences between the WMS and control cases. The independent measures included diagnosis (WMS and control), hemisphere (right and left), and layer (II, III, IVA, IVB, IVCα, IVCβ, V, and VI). The dependent measures were cell size (areas of the nucleus and cytoplasm together) and cell-packing density. The effect of sex could not be analyzed with any confidence because of the few cases studied. Differences in neuronal size distributions were analyzed using χ² tests.

CELL-PACKING DENSITY

Repeated-measures ANOVA revealed significant differences in cell-packing density between the WMS-affected cases and controls and between hemispheres. As expected, based on the known difference in neuronal types among the layers, there was a significant effect of layer overall (F7,28=23.28, P<.001) and also for the left hemisphere (F7,28=27.42, P<.001) and the right hemisphere (F7,28=9.67, P<.001) separately. A hemisphere by layer interaction was significant only when layer IV (all sublaminae combined) was analyzed (F3,12=3.58, P<.05). Individual analyses of layers III and IV showed a significant increase in cell-packing density in the left hemisphere in the WMS-affected brains in sublayer IVcβ (F1,4=8.35, P<.05) compared with the controls (309598 vs 210526 neurons/mm³) (Figure 3) but not in layer III. There was also a significant interaction between layer and diagnosis in the left hemisphere for layer IV (F3,12=3.58, P<.05), but not in the right hemisphere.

MEAN NEURONAL SIZE

Repeated-measures ANOVA analyses of cross-sectional mean neuronal areas did not result in any significant differences between WMS-affected and control brains. As expected, by the known differences in cell size between layers, there was a significant main effect of layer over both hemispheres (F7,28=20.63, P<.001) and for each hemisphere analyzed separately: left (F7,28=23.38, P<.001) and right (F7,28=11.11, P<.001).

NEURONAL SIZE DISTRIBUTIONS

Where there is marked variability in neuronal size, as is the case in the cerebral cortex, significant neuronal differences may be difficult to demonstrate, as in the case of...
where both large and small neurons increase in numbers. Therefore, to assess additional differences in neuronal size in each layer and sublayer, we analyzed the frequency distribution of cell size in consecutive bins (Figure 4). The bins were arranged in ascending order of cell size. The number of bins ranged from 7 to 12, increasing by 10 µm², and contained neurons whose size ranged from 30 to 90 µm². We calculated χ² Values for the distribution of neurons in these bins between WMS-affected and control brains. We also examined distribution of cell size differences between hemispheres for WMS-affected and control brains. We further observed that the distribution of cell size in consecutive bins between WMS-affected or control brains.

In this study, we sought an anatomical explanation for the unique behavioral profile of individuals with WMS. For example, some behaviors are deeply abnormal while others are relatively preserved. Among the relatively preserved behaviors, we find language that is rich in vocabulary and affective prosody as well as excellent face recognition abilities and verbal memory. More severely affected behaviors include visuospatial-visuomotor abilities and mathematics. As shown in Figure 1, individuals with WMS show specific deficits in spatial cognition tasks such as object assembly, block design, and drawing, suggesting difficulty with overall configurational task performance. More involvement of visuospatial functions implicates either the right hemisphere or the dorsal visual pathway, whereas visual object recognition involves the left hemisphere or the ventral visual pathway. In WMS-affected subjects, therefore, preservation of facial recognition abilities and affective speech prosody, which are linked to right hemisphere function, makes us suspect that the visuospatial and visuomotor deficits are not explained by a right hemisphere problem, but rather by a problem with the dorsal visual pathway and its connections to the motor system. The present study was designed to test this hypothesis.

Based on anatomical, physiological, and clinical data, our hypothesis was that the abnormalities in WMS would be found in visual cortex that projects to the dorsal system (the part representing peripheral fields in the ante-
rior calcarine region), would affect neurons that form part of the magnocellular system, and would be more striking in the right hemisphere. The anterior calcarine cortex was sampled and, in fact, the findings were nearly the opposite. Specifically, although the peripheral visual cortex was found to be abnormal in WMS-affected brains, parvocellular sublayers in the left hemisphere only were involved.

In the primate visual system, there are structural and functional distinctions between 2 relatively segregated and independent processing pathways—the parvocellular and the magnocellular systems. These pathways have been characterized on the basis of anatomical,41 psychophysical,42,43 and physiological properties.28,44 Neurons in these 2 systems differ in terms of receptive field size, sensitivity to color and light contrast, and timing properties. The parvo system is ideally suited for form, texture, and color analysis, while magno processes larger sections of space and appears better designed to calculate spatial location and motion. Anatomically, the magnonurons are restricted to the lower 2 layers of the lateral geniculate nucleus, whereas parvo cells occupy the upper 4 layers. In the cortex, although a separation still exists (see below), it is less absolute.12,45 For example, evidence from primate work suggests that segregation of magnocellular and parvo-cellular signals continue into extrastriate visual areas into higher-order visual processing. On the other hand, some primate studies suggest that magnocellular and parvocellular streams contribute differentially to dorsal and ventral pathways46 and that V1 neurons integrate some information carried by both lateral geniculate nucleus magnocellular and parvocellular pathways.47 Similarly, the visual system is subdivided into a dorsal and ventral pathway based on anatomical location and behavioral studies in monkeys and humans.11,14,46,48 The relationship between the parvo-magno and dorsal-ventral subdivisions remains tentative49 and also controversial,12 but one may argue that the magnocellular system is the one more likely to contribute particularly to the dorsal visual system, with the parvo system contributing more specifically to the ventral system.50,51 Also, based on clinical findings and activation studies, one would then be able to suggest that the magnocellular system is not only dorsal but also lateralized to the right hemisphere, with the parvo system being lateralized to the left.15,30,48 This suggestion is based on clinical observations that right hemisphere lesions tend to affect visuospatial abilities while left hemisphere lesions evince more clearly as visual object anomalies and agnosias.52 The findings of this study show that neuronal differences between WMS-affected cases and controls in primary visual cortex appear to affect the left hemisphere more than the right, particularly layer IV. Furthermore, the most consistent finding emerged in cell size distribution: WMS-affected visual cortex had less large and fewer large neurons than the control brains in layers IVA and IVCβ. This layer receives inputs from the lateral geniculate nucleus. The lateral geniculate nucleus inputs to layer IV are sub-

CONCLUSIONS

These conclusions are based on the assumption that the sampled area represents the peripheral visual fields in the WMS-affected brains and, thus, the dorsal visual pathways. We can be certain of this in the control brains, but less so in the WMS-affected cases. Thus, even though re-
cent maps of central vision show an expanded area sub-servicing foveal vision than earlier estimates, we are certain to have sampled peripheral visual cortex in controls. However, if it were the case that central vision is even more expanded in WMS (which makes sense on theoretical grounds), it could be that, while we thought we were sampling peripheral visual cortex on topographic grounds, we were actually sampling cortex representing central vision in the WMS-affected cases. In that case the comparisons to the control samples would not be meaningful. While we cannot solve this conundrum—to ascertain the functional topography of the visual cortex—until functional activation studies are performed in WMS-affected subjects, our postmortem findings—more compaction and smaller cells in WMS-affected brains—may be related to the visuospatial deficits in this intriguing syndrome.

Accepted for publication March 7, 2002.

Author contributions: Study concept and design (Drs Galaburda and Holinger); acquisition of data (Drs Galaburda, Holinger, Bellugi, and Sherman); analysis and interpretation of data (Drs Galaburda, Holinger, Bellugi, and Sherman); drafting of the manuscript (Drs Galaburda, Bellugi, and Sherman); critical revision of the manuscript for important intellectual content (Drs Galaburda, Holinger, and Sherman); statistical expertise (Drs Holinger and Sherman); obtained funding (Drs Galaburda and Bellugi); administrative, technical, and material support (Dr Galaburda); study supervision (Dr Galaburda); graphics (Dr Holinger).

The study was supported by grant HD33113 to the Salk Institute for Biological Studies from the National Institutes of Health, Bethesda, Md (Dr Bellugi). The brain tissue for controls was provided by the Harvard Brain Tissue Resource Center supported in part by grant MN/NS 31862 from the Public Health Service, Washington, DC.

We are grateful to the families who have donated the WMS-affected brains and to the local, regional, and national Williams Syndrome Associations. We appreciate technical assistance from Antis Zalkalns, BS.

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REFERENCES


44. Allison JD, Meltzer P, Ding Y, Bonds AB, Casagrande VA. Differential contributions of magnocellular and parvocellular pathways to the contrast response of neurons in bush baby primary visual cortex (V1). Vis Neurosci. 2000;17: 71-76.